



# Assessment the Effect of *Ficus carica* Leaf Extract on B<sub>16</sub>F<sub>10</sub> Melanoma Cancer Cells: The Roles of p53, Caspase-3 & Caspase-9 on Induction of Intrinsic Apoptosis Cascade

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## Abstract

**Background:** Melanoma is the most serious kind of skin cancer which has significantly increased in recent decades, and the importance of its primary treatment is increased widely. *Ficus carica* leaves have various therapeutic impact such as anti-inflammatory, anti-proliferative and apoptotic activity.

**Objectives:** Hence, regarding the *F. carica* effect on the treatment of various diseases, the present research was conducted to identify the effect of methanolic extract of *F. carica* leaf on apoptosis induction in B<sub>16</sub>F<sub>10</sub> melanoma cancer cells.

**Methods:** Cell survival was estimated by MTT assay after treatment of B<sub>16</sub>F<sub>10</sub> cells in various concentrations of *F. carica* leaf extract (150, 250, 350, 450, 550, 650, 750 and 850 µg/mL) for 24 and 48 h. Cell apoptosis was analysed by AO/PI and DAPI staining, Annexin V/Propidium Iodide flow cytometry. Moreover, Real-time PCR was utilized to evaluate the expression of apoptotic genes including p53, Bax, caspase-3 and caspase-9 genes.

**Results:** MTT assay results indicated that methanolic extract of *F. carica* leaf prevented the proliferation of B<sub>16</sub>F<sub>10</sub> cells in a dose and time dependent manner. AO/PI staining results showed an elevation in apoptotic cells in treated groups and DAPI indicated that *F. carica* extract resulted in chromatin condensation and fragmentation. Annexin V revealed the increasing percentage of apoptotic cells after treatment. In addition, the up-regulation of apoptotic genes confirmed the apoptosis inducing potential of *F. carica* leaves in B<sub>16</sub>F<sub>10</sub> cells by Real-time PCR.

**Conclusions:** Thus, methanolic extract of *F. carica* leaves could be suggested as an effective anti-cancer agent for further studies on melanoma cancer.

**Keywords:** *Ficus carica*, Melanoma Cancer, Anti-proliferative, Apoptosis, Cytotoxicity

## 1. Background

Cancer is regarded as a multi-stage process in which the genetic and epigenetic changes in a cell, result in a progressive alteration from normal to cancer cells (1). Several factors are involved in developing cancer (2), the internal reasons of cancer may be related to oxidative stress generation, apoptotic process disorders, and genetic mutations; hence the external reasons of cancer may be consistent with the environmental factors like viruses, bacteria, smoking, chemicals, radiation, and stress (3). Several researchers indicated that oncogenes activation and the inaction of tumor suppressors are needed for cancer cells proliferation (4). The main genes lead to cancer are included in apoptosis, tumor suppressors, onco-

genes, and DNA repair genes (1). Apoptosis is a kind of gene-dependent programmed cell death identified by special morphological and biochemical properties, like chromatin density, cell shrinkage, and nuclear condensation (5). The cell death process can be induced by extracellular endogenous proteins or cytokines and oxidative stress reactions (6). In apoptotic process, caspases are among the most significant factors involved in apoptosis (7). The name of caspases is derived from their function and these enzymes coordinate the route of programmed cell death (6). Skin cancer (melanoma) is a common type of cancer, and early detection increases survival (8). Conventional treatments for melanoma involve surgery, chemotherapy, and immunotherapy (9). Nowadays, general tendency of society to utilize herbal medicines and natural products

is raising, so that herbal medicines have an important effect on pharmaceutical products (10). *Ficus carica* (fig) belongs to Moraceae family, the name of species derived from Carica is the name of fig-growing region in the southwest of Turkey. Its major origin is Mediterranean region and it grows in multiple regions of the world (11). Figs have an essential role to reduce cholesterol, controlling respiration and strengthening the heart. Other features involve decreasing body temperature, increasing body sweating, and treating anemia (12). Fig leaf extract has anti-cancer, anti-proliferative effects (13). The most significant compounds in figs are proteins, fatty acids, enzymes, beta-carotene, fucosin, arabinose, and xanthotoxols (14). Furthermore, it involves phenolic compounds like catechin, gallic acid and syringic acid (15). *Ficus carica* leaf extract was indicated to involve rhein, chrysin and quercetin which can prevent proliferation of cancer cells, and rhein has anticancer impacts (16). *Ficus carica* leaf extract has a stimulating impact on cellular and humoral immune response; hence it can be applied for therapeutic objectives in the diseases of immune origin (13).

## 2. Objectives

Regarding different chemical compounds in *F. carica* and therapeutic features of this plant to treat various diseases, we decided to survey the impact of *F. carica* leaves extract on apoptosis induction in melanoma cancer cells (B<sub>16</sub>F<sub>10</sub>).

## 3. Methods

### 3.1. Chemicals

B<sub>16</sub>F<sub>10</sub> cell line (melanoma cancer) was obtained from Pasteur Institute of Iran. Culture medium DMEM (Dulbecco's Modified Eagle Medium), FBS (Fetal bovine serum), antibiotics (penicillin-streptomycin) and Trypsin-EDTA were provided from Bioidea (Iran). MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide), DAPI (4',6-Diamidino-2-phenylindole dihydrochloride), Propidium Iodide and Acridine orange were provided from Sigma (Germany), Annexin V-FITC kit was provided from Abcam (UK), as well as SYBR Green, RNA extraction kit and CDNA synthesis were provided from Pars Tous (Iran).

### 3.2. Plant Materials

*Ficus carica* leaves were provided from Mashhad (Khorasan Razavi, Iran) in May 2019, and confirmed in Islamic Azad University, Mashhad botanical lab with herbarium code 10319.

### 3.3. Extraction Method

In this study, the dried *F. carica* leaves were crushed into a powder and 100cc of methanol per 1 gram was added to it. Then, the resultant mixture was kept in the dark at room temperature for 72 h. During this time, the mixture was well stirred three times a day and the extract was filtered using Whatman paper after 72 h, and the total methanolic extraction was performed by using rotary evaporator. To evaporate methanol, the resulting extract, which was water-insoluble green precipitate was placed at 66.5°C for 72 h.

### 3.4. Cancer Cell Survival Analysis

Melanoma cancer cells (B<sub>16</sub>F<sub>10</sub>) and normal epithelial kidney cells (HEK-293) were cultured in DMEM medium supplemented with 10% FBS and 1% antibiotic. The cells were maintained at 37°C in a 5% CO<sub>2</sub> atmosphere and 95% humidity. Cell viability was examined by MTT assay. Briefly, B<sub>16</sub>F<sub>10</sub> and HEK-293 cells were cultured in 96-well plate with  $1 \times 10^4$  per well and incubated for 24 hours. Cells were treated by different concentrations of the methanolic extract of *F. carica* leaf (150, 250, 350, 450, 550, 650, 750 and 850 µg/mL) and sham (DMSO as extract diluent) for 24 and 48 hours. After incubation, MTT solution was added to culture medium which was maintained in darkness for 4 hours at 37°C. Next, the culture medium was discarded and 80 µL of DMSO was added to cells. The absorbance was estimated at 560 nm. Then, the results were compared to control groups.

### 3.5. DAPI Staining

In this study, the nuclear morphology alteration was assessed with DAPI dye. B<sub>16</sub>F<sub>10</sub> cells were treated in various concentrations of 350, 450, 550, 650 and 750 µg/mL of *F. carica* leaves extract which incubated for 24 hours. Then, the cells were fixed and stained with methanol and DAPI, respectively. Nucleus morphology was seen under fluorescence microscope (Olympus, Japan) via blue filter, the excitation and emission wavelength were 364 nm and 454 nm respectively.

### 3.6. Acridine Orange-Propidium Iodide (AO/PI) Staining

Cell viability and apoptosis were assessed by fluorescence microscopy utilizing AO/PI. In brief, B<sub>16</sub>F<sub>10</sub> cells were incubated for 24 h at 350, 450, 550, 650 and 750 µg/mL concentration of methanolic extract of *F. carica* leaves. Then, the cells were trypsinized and centrifuged. The cells were stained with 10 µL of a mixture of AO and PI fluorescence. At last, the cells were seen under ultraviolet fluorescence microscope (Olympus, Japan) at 490 nm excitation and 520 nm emission wavelength.

### 3.7. Annexin V Fluorescence Assay

In this experiment, apoptosis and necrosis cells were quantified by flow cytometry. In brief, melanoma cells were treated by multiple concentrations of *F. carica* leaves extract. After 24 hours, the cells were collected and 500  $\mu$ L of binding buffer 1X was added to every sample. Subsequently, 5  $\mu$ L of Annexin V-FITC and 5  $\mu$ L of propidium iodide were added to every sample which was estimated by FACSCalibur flow cytometer (BD Biosciences).

### 3.8. Real-Time Quantitative Polymerase Chain Reaction

The expression levels of Bax, caspase3, caspase9, and p53 genes were assessed by real-time PCR. The total RNA was extracted from melanoma cancer cells after treatment with various concentrations of *F. carica* leaf extract (350, 450, 550, 650 and 750  $\mu$ g/mL), due to the manufacturer's instructions. The extracted RNA was utilized for cDNA synthesis regarding Pars Tous manufacturer instructions. The specific primer sequences for p53, Bax, caspase3, caspase9 and GAPDH genes was designed as shown in Table 1. The relative quantification was performed in real-time PCR detection system (Bio-Rad CFX96) using SYBR Green. The results were normalized to an internal reference gene GAPDH. All PCR amplifications were analyzed in triplicates.

**Table 1.** Primer Sequences of Genes

Gene	Sense	Anti-sense
GAPDH	AATCCCACTCTTCCACCTTCG	GTCCACCACTCTTGTCTGTAG
p53	AACCGCCGACCTATCCTTACC	TCCAGGGCGGCACAAAC
Bax	TCATGGGCTGGACACTGGACTTC	GAGCGAGGCGGTGAGGACTC
Caspase3	TGACTGGAAGCCGCCGAACTC	GCAAAGCCATCTCTCATCAG
Caspase9	GAAGAACGACCTGACTGCCAAG	GAGAGAGGATGACCACCACAAAG

### 3.9. Statistical Analysis

All results were expressed as mean  $\pm$  SEM. Statistical analysis was performed using GraphPad Prism 8 software, ANOVA test followed by Tukey test to compare the data and  $P < 0.05$  level was considered significant.

## 4. Results

### 4.1. Cancer Cell Survival Analysis

The result of MTT assay showed that *F. carica* leaves extract had cytotoxic impacts on B<sub>16</sub>F<sub>10</sub> cells in a dose- and time-dependent manner as compared with control or

Sham (DMSO) groups (Table 2). Therefore, our data indicated the inhibitory impact of *F. carica* extract on skin cancer cells by increasing extract concentration. Due to our results, the concentration of 750 and 650  $\mu$ g/mL of methanolic extract of *F. carica* leaves decreased cells viability (IC<sub>50</sub>) by 50% for 24 h and 48 h respectively. However, these concentrations did not exert high cytotoxic activity against normal epithelial HEK-293 cells and indicated that B<sub>16</sub>F<sub>10</sub> cancer cells were more sensitive to *F. carica* leaves extract than normal cells (Table. 2).

### 4.2. DAPI Staining

Regarding DAPI staining results, increasing concentration of the methanolic extract of *F. carica* leaves result in condensed nuclei and their DNA was seen as fragmented that confirmed cells underwent apoptosis. The apoptosis population was significantly higher after treatment with 650 and 750  $\mu$ g/mL of extract (Figure 1). While in control group, the stained nuclei of B<sub>16</sub>F<sub>10</sub> cells maintained the round shape and homogeneous.

### 4.3. AO/PI Staining

Staining with AO/PI indicated that *F. carica* treatment promoted apoptosis in melanoma cancer cells (B<sub>16</sub>F<sub>10</sub>). Figure 2 showed control cells were stained green with intact nucleus structure while the apoptosis was clearly obvious in cells treated with cytotoxic concentrations of methanolic extract of *F. carica* leaf as marked by orange nuclei and red staining represents necrosis.

### 4.4. Annexin V-FITC Test

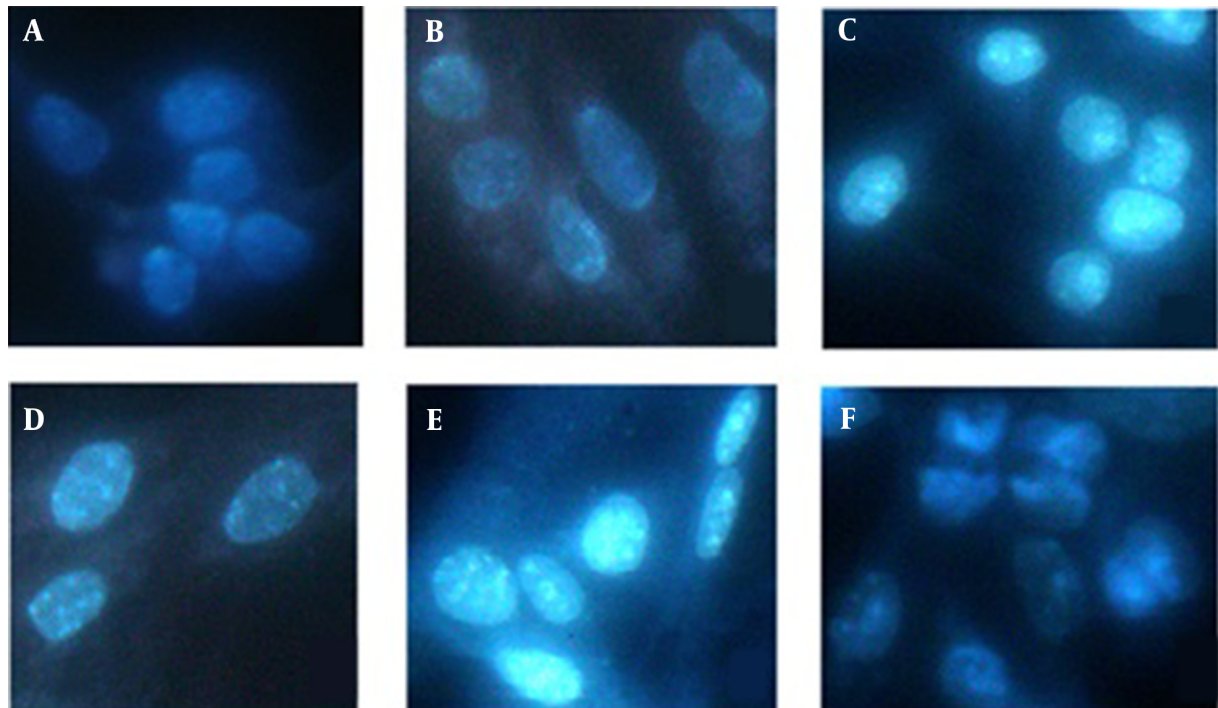
Detection of apoptosis by Flow cytometry revealed that the percentage of apoptotic cells increased after methanolic extract of *F. carica* leaves treatment in a dose-dependent manner. Due to flow cytometry histogram, the lower concentrations of *F. carica* leaf extract, exhibited the least population of apoptosis and higher viable cell. In contrast, the percentage of early and late apoptosis increased in higher concentrations of *F. carica* extract (Figure 3 and Table 3).

### 4.5. Real-time PCR

The alteration in expression of p53, Bax, caspase 3 and caspase 9 genes in melanoma cancer cells treated at concentrations of 350 to 750  $\mu$ g/mL of *F. carica* leaves extract were evaluated by real-time PCR. Our data revealed that increasing expression of p53, Bax, caspase 3 and caspase 9 genes in treated group compared to control which confirmed the apoptosis induction in B<sub>16</sub>F<sub>10</sub> cells through the activation of mitochondrial pathways (Figure 4). By increasing concentrations of *F. carica* leaf extract, particularly at high concentrations from 650 to 750  $\mu$ g/mL, genes expression involved in apoptosis has increased than reference gene in skin cancer cell line.

**Table 2.** Effect of *Ficus carica* Leaves Extract on Cell Viability of B<sub>16</sub>F<sub>10</sub> and HEK-293 Cells <sup>a</sup>

Groups	HEK-293, 24 h	B <sub>16</sub> F <sub>10</sub> , 24 h	B <sub>16</sub> F <sub>10</sub> , 48 h
Control	100.00 ± 1.13	100.00 ± 1.55	100.00 ± 1.23
Sham	98.16 ± 1.23	98.03 ± 1.63	98.31 ± 2.20
F.carica leaves 150 µg/mL	97.10 ± 1.18	97.42 ± 2.23	94.78 ± 2.94
F.carica leaves 250 µg/mL	97.55 ± 0.74	95.21 ± 1.35	90.43 ± 5.04
F.carica leaves 350 µg/mL	95.95 ± 2.24	90.47 ± 5.02	85.42 ± 2.92 <sup>b</sup>
F.carica leaves 450 µg/mL	91.88 ± 0.39	82.39 ± 2.71 <sup>c</sup>	78.36 ± 2.72 <sup>c</sup>
F.carica leaves 550 µg/mL	87.47 ± 1.49 <sup>b</sup>	69.17 ± 4.70 <sup>c</sup>	62.82 ± 3.35 <sup>c</sup>
F.carica leaves 650 µg/mL	75.37 ± 1.86 <sup>c</sup>	54.38 ± 3.28 <sup>c</sup>	49.00 ± 3.11 <sup>c</sup>
F.carica leaves 750 µg/mL	62.98 ± 1.66 <sup>c</sup>	45.61 ± 4.60 <sup>c</sup>	41.87 ± 3.23 <sup>c</sup>
F.carica leaves 850 µg/mL	58.71 ± 1.06 <sup>c</sup>	41.23 ± 3.50 <sup>c</sup>	34.84 ± 5.13 <sup>c</sup>

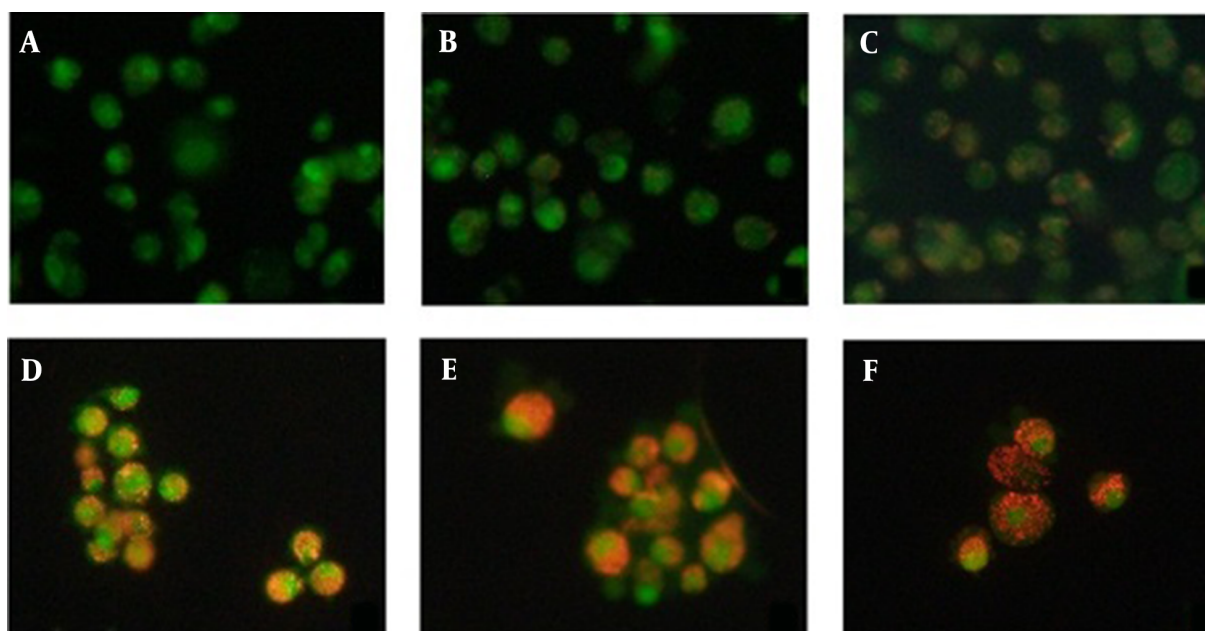
<sup>a</sup> Data are represented as the mean ± SD.<sup>b</sup> P value significant at the level of P < 0.01 compared to the control.<sup>c</sup> P value significant at the level of P < 0.001 compared to the control.**Figure 1.** DAPI staining of untreated and treated B<sub>16</sub>F<sub>10</sub> cells. (A) Control, (B-F) concentration of 350, 450, 550, 650 and 750 µg/mL of *Ficus carica* leaves extract (magnification X400).

## 5. Discussion

Epidemiological studies indicate the significant prevalence of skin cancer and mortality in recent years (17). Melanoma, as serious skin cancer, develops in melanocytes. Melanocytes make melanin, or skin pigment. The strongest risk factors for melanoma are differ-

ent abnormal moles and a family history of melanoma (18). Apoptosis is a very regular and precise process which is regulated by a series of signaling cascades and cellular proteins. Thus, a deep understanding of apoptosis mechanisms and related pathways is efficient not only in understanding the disease but also in disease treatment (3). Apoptosis defects play a significant role in tumor





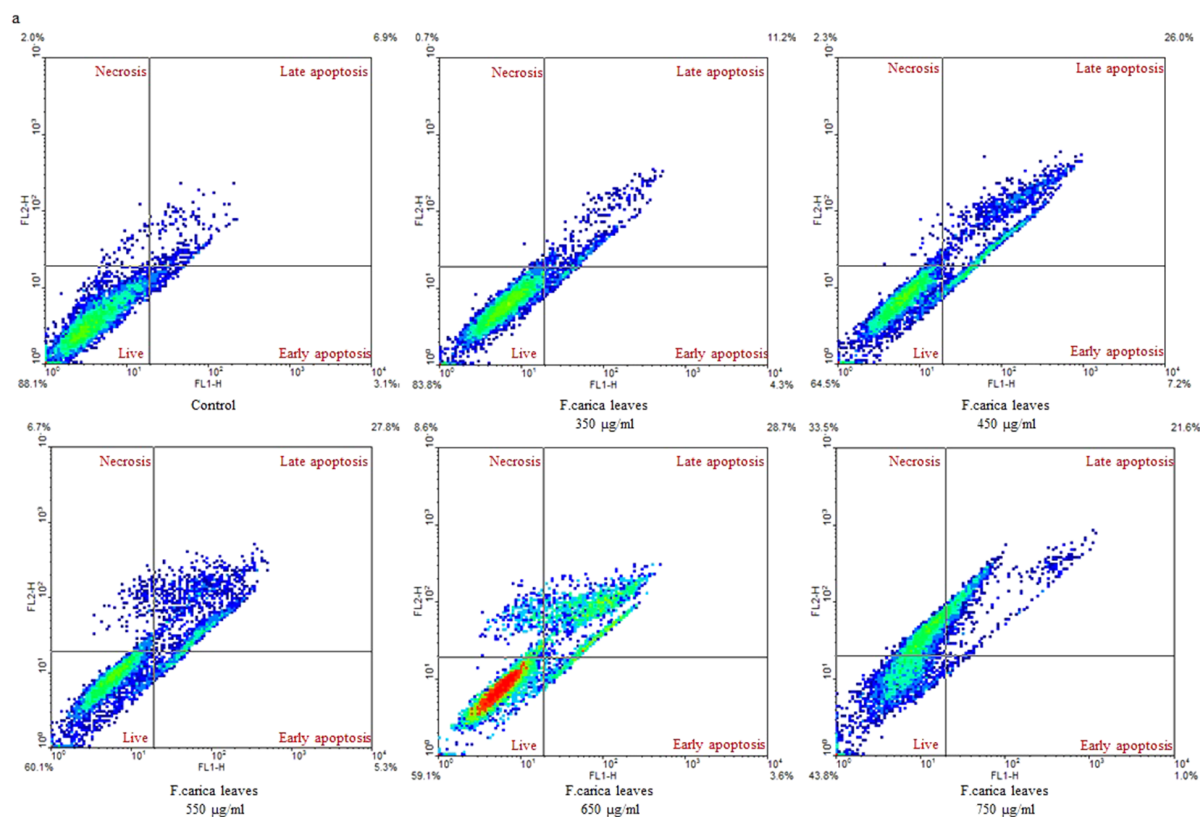
**Figure 2.** Acridine orange/puoropidium iodide staining. (A) Control, (B-F) concentration of 350, 450, 550, 650 and 750  $\mu\text{g/mL}$  of *Ficus carica* leaves extract (magnification X100).

**Table 3.** The Percentage of Apoptotic Cells Indicating the Effect of *Ficus carica* Leaves Extract on Apoptosis in  $\text{B}_{16}\text{F}_{10}$  Cells

Group	Liver Cells (%)	Early Apoptotic Cells (%)	Late Apoptotic Cells (%)	Necrotic Cells (%)
Control	88.1	3.1	6.9	2.0
<i>Ficus carica</i> 350 $\mu\text{g/mL}$	83.8	4.3	11.2	0.7
<i>F. carica</i> 450 $\mu\text{g/mL}$	64.5	7.2	26.0	2.3
<i>F. carica</i> 550 $\mu\text{g/mL}$	60.1	5.3	27.8	6.7
<i>F. carica</i> 650 $\mu\text{g/mL}$	59.1	3.6	28.7	8.6
<i>F. carica</i> 750 $\mu\text{g/mL}$	43.8	1.0	21.6	33.5

formation and regulation disruption causes resistance to chemotherapy and radiation therapy. Moreover, this trend may increase metastasis. Any factor which can inhibit normal development of cells provides conditions for apoptosis. Thus excessive apoptosis can result in diseases like Parkinson's and Alzheimer's and ischemic injuries (19). *Ficus carica* have been utilized in conventional medicine to treat numerous diseases including cancer. Some of the phenolic compounds in fig leaves include furanocoumarins such as psoralen and bergapten, flavonoids such as quercetin 3-O rutinoides, and phenolic acids such as ferulic acid, 3-O-caffeic acid and 5-O-caffeic acid (20). Furanocoumarin and psoralen show the anticancer and anti-cholinesterase activities, and flavonoids indicate anticancer, anti-inflammatory, and antidiabetic activities (13). Purushotham et al. investigated the anti-proliferative, anti-inflammatory and anti-apoptotic activity of methano-

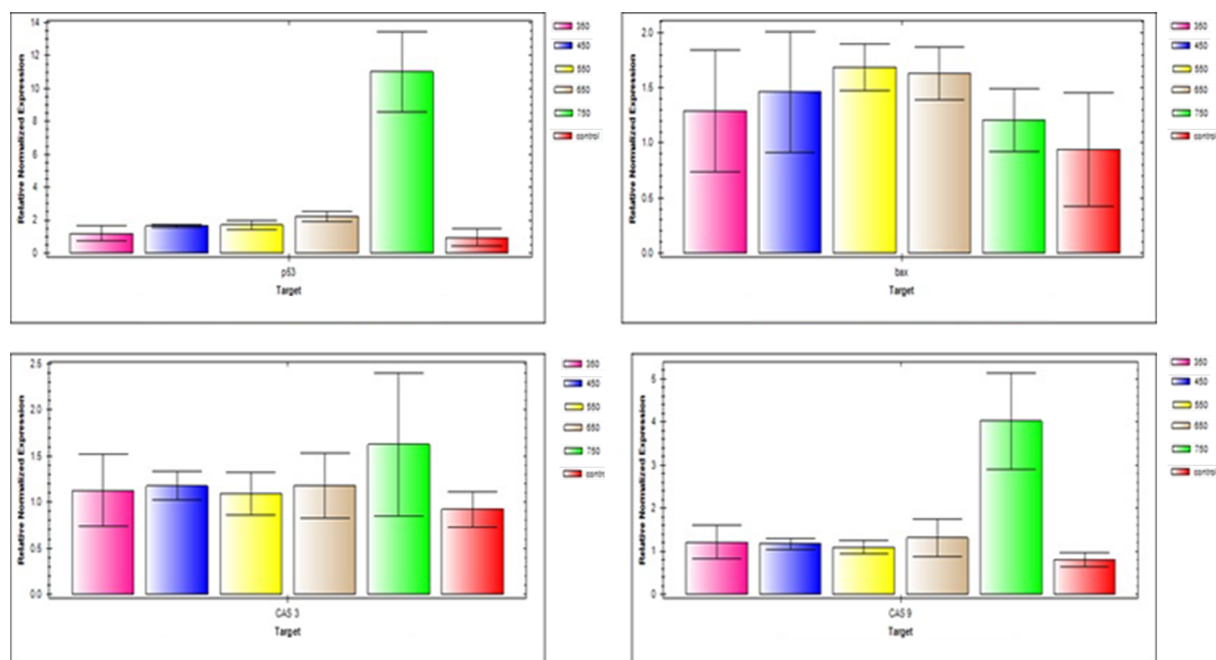
lic extract of *Andrographis paniculata* (AN) in A375 and  $\text{B}_{16}\text{F}_{10}$  cell lines. Due to their results, methanolic extract of AN leaf and stem caused a decrease in  $\text{B}_{16}\text{F}_{10}$  cell viability. In addition, leaf extract had a higher anticancer activity compared with its stem on cancer cells. These results revealed that AN decreased the proliferation and induced apoptosis in cancer cell lines (21). Regarding obtained results, increasing the dose of methanolic extract of *F. carica* leaf induced apoptosis in  $\text{B}_{16}\text{F}_{10}$  cancer cells, followed by preventing cancer cell proliferation. Goldberg et al. (2017) surveyed the impact aqueous extract of *Hibiscus rosa sinensis* compounds in vitro. The results confirmed that extract contained compounds which prevented the growth of melanoma cells by dose-dependent manner (22). Our results including MTT, Annexin V/ PI, DAPI, and AO/PI assays all indicated that  $\text{B}_{16}\text{F}_{10}$  cells treated with methanolic extract of *F. carica* leaves extract promote



**Figure 3.** Annexin V-FITC test results. (A) Flow cytometry plots, (B) The percentage of apoptotic cells indicating the effect of *Ficus carica* leaves extract on apoptosis in B<sub>16</sub>F<sub>10</sub> cells.

the apoptosis through the activation of mitochondrial pathways. Regarding increased expression of genes involved in mitochondrial pathways of caspase 3 and caspase 9, methanolic extract of *F. carica* leaves induces apoptosis in B<sub>16</sub>F<sub>10</sub> cells. methanolic extract of *F. carica* leaf prevent proliferation of skin cancer cells (B<sub>16</sub>F<sub>10</sub> cell line) which the results are consistent with above research. MTT test results indicated that *F. carica* leaves extract has a cytotoxic impact on B<sub>16</sub>F<sub>10</sub> cell line in dose and time dependent manner and decreased the cells viability and induced apoptosis. It is followed by inhibiting melanoma cancer cell proliferation. Zhang et al. investigated the inhibition of pancreatic  $\beta$ -cell apoptosis by the lower concentration of *F. carica* leaves extract (FCL) through the AMPK/JNK/Caspase 3 signaling pathway by antioxidation properties. They demonstrated that FCL at 60  $\mu$ g/mL concentration could inhibit apoptosis induced by hydrogen peroxide and reduce ROS production. Therefore, FCL could inhibit pancreatic  $\beta$ -cell apoptosis by inhibiting the AMPK/JNK/ Caspase 3 signaling pathway and by antioxidation properties at lower concentration of FCL (60

$\mu$ g/mL) (23). In the present study, the results of Annexin, MTT, DAPI tests indicated that in B<sub>16</sub>F<sub>10</sub> cells treated with the methanolic extract of fig leaves at lower concentration have no apoptotic and inhibiting effect on B<sub>16</sub>F<sub>10</sub> cells while the higher concentration of *F. carica* leaves extract (650  $\mu$ g/mL) induced apoptosis and activated mitochondrial pathways, which inhibited the proliferation of the skin cancer cells. Turkoglu et al. investigated the effect of extract of *F. carica* leaf on the expression of inflammatory factor genes in HaCaT cells. The results of gene expression analysis by Real time PCR showed that extract of *F. carica* leaf significantly reduced interleukin-1, TNF- $\alpha$  compared to the untreated cells. Moreover, based on the results of cytotoxicity analysis, the extract of *F. carica* leaf reduced the proliferation of HaCaT cells. Therefore, this extract is useful for some inflammatory and skin disorders such as androgenic alopecia (24). In addition, the MTT test showed that the alcoholic extract of fig leaves had a cytotoxic effect on B<sub>16</sub>F<sub>10</sub> cells, which reduced the viability of cells in low concentrations. Thus, the cells underwent apoptosis, which caused the inhibition of the proliferation of



**Figure 4.** Expression pattern of p53, Bax, caspase3, and caspase9 genes in melanoma cancer cells treated at different concentrations of 350, 450, 550, 650 and 750 µg/mL.

melanoma cancer cells. In 2018 Zhang et al. analyzed the extracts and compounds of *F. carica* leaves on cell survival, cell cycle and breast cancer cell migration (MDA - MB-231 cell line), and consistent with our results their results indicated that chemical compounds in *F. carica* leaves like Psoralen and Bergapten caused a decrease in breast cancer cell proliferation with a time and dose-dependent effect. Therefore, these extracts and compounds prevent survival of cells and the cell cycle of cancer cell line (25). Purnamasari et al. evaluated the anticancer activity of the alcoholic extract of *F. carica* leaf and fruit against proliferation, apoptosis and necrosis in Huh7it cells. Based on the results, *F. carica* is a rich source of polyphenols, and the results of MTT and AnnexinV assays indicated that the difference in anticancer activity is related to the various compounds in each extract (26). In this regard, the results of the present study indicated that the methanolic extract of fig leaves inhibits the proliferation of skin cancer cells.

### 5.1. Conclusions

Using herbal and traditional remedies in medicine and pharmacy opened a novel insight for researchers to treat various disease. Our results indicated the efficiency of *F. carica* leaves extract on preventing proliferation and apoptosis induction in melanoma cancer cells. It is expected that in near future, the impact of *F. carica* leaves, its efficient compounds and related signaling pathways which

induce apoptosis in cancer cells will be addressed.

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### Footnotes

**Authors' Contribution:** Study concept and design: J. B.; Analysis and interpretation of data: J. B., M. S., and F. S.; Drafting of the manuscript: J. B., M. S., and F. S.; Critical revision of the manuscript for important intellectual content: J. B., F. S., and E. A.; Statistical analysis: J. B., E. A., and F. S.

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