






Impact of Berberine on the Expression of Apollon Gene in Chronic Lymphocytic Leukemia

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Abstract

Background: The incidence of B-chronic lymphocytic leukemia (B-CLL) resulting from the clonal accumulation of apoptosis-resistant malignant B lymphocytes is growing in the adult population of Iran. Inhibitors of apoptosis proteins (IAPs) are considered as factors that can delay the onset of CLL cell apoptosis. Berberine is an isoquinoline alkaloid isolated from *Cotridis rhizoma* that exhibits anti-tumor activities through various mechanisms.

Objectives: In this study, we investigated the impact of berberine on the level of Apollon expression in peripheral blood mononuclear cells (PBMCs) of 12 cases newly diagnosed with CLL and 6 healthy donors.

Methods: At first, the level of Apollon expression was assessed in PBMCs of CLL patients compared to the healthy donors. Peripheral blood mononuclear cells were cultured in RPMI-1640 medium with 5% fetal bovine serum (FBS) and 1% penicillin/streptomycin for 48 hours, and the effect of berberine (25 μ M) on the level of Apollon expression in CLL patients was assessed and compared to that of healthy donors.

Results: We found that the expression level of Apollon was not significantly different between CLL patients and healthy donors ($P = 0.640$). Moreover, berberine induced no significant differences in Apollon expression as compared to the untreated (control) group ($P = 0.545$ and $P = 0.267$ in CLL patients and healthy donors, respectively).

Conclusions: Overall, our results suggest that berberine has no direct effect on the expression of Apollon gene in CLL patients, and pro-apoptotic impacts of berberine may be exerted through other mechanisms.

Keywords: B-Cell Chronic Lymphocytic Leukemia, Apoptosis, Berberine, Apollon Protein

1. Background

B-chronic lymphocytic leukemia (B-CLL) is characterized by the abnormal accumulation of B lymphocytes in the peripheral blood, bone marrow, and lymphoid tissues. Chronic lymphocytic leukemia is the most prevalent adult leukemia in Western countries and accounts for 11 - 15% of total hematopoietic cancers in Iran (1). The incidence rate of this condition increases in adults with median age of 72 years. Moreover, males are more affected than females (1.7: 1) (2). Morphologically, CLL cells are considered as long-lived, small, mature, and resting lymphocytes with the CD5+, CD19+, and CD23+ phenotypes (3).

However, alteration in the capacity of cell apoptosis that leads to resistance to anti-tumor immune response is a principal feature of CLL (4). There is strong consensus that defect in apoptosis regulation leads to the accumulation of malignant B cells. Apoptosis as programmed cell death is executed through the activity of caspase fam-

ily and controlled by several checkpoints (5). One of the apoptosis regulation checkpoints is the control of caspase activation through the inhibitor of apoptosis protein (IAP) family, including XIAP, cIAP1, cIAP2, ILP-2, NAIP, survivin, and BRUCE/Apollon. The Inhibitors of apoptosis protein consist of at least one baculovirus IAP repeat (BIR) domain, C-terminal RING zinc-finger domain, and caspase recruitment domain (CARD) (6). Moreover, IAPs play a central role in the cell cycle and signal transduction.

It is demonstrated that IAPs are tightly associated with RE1-silencing transcription factor (REST) expression, as a master regulator in tumor development, progression, and metastasis (7). Therefore, it seems that the dysregulation of IAPs may lead to tumorigenesis. In this context, Apollon/BIRC6 (BIR-containing protein 6) has been identified as a 530kDa protein that its encoding gene is located on the chromosome band 2p22. It is demonstrated that Apollon exerts its anti-apoptotic activity through promoting ubiq-

uitation, and subsequently, the degradation of the pro-apoptotic protein Smac/DIABLO. It is also able to bind to pro-caspase-9 and inhibit its activation (8). Accordingly, the overexpression of BIRC6 has been demonstrated in hematological malignancies, including childhood acute myeloid leukemia (AML), acute lymphoblastic leukemia (ALL), and CLL, and it has been associated with poor prognosis and chemoresistance (9, 10).

In addition to the reported beneficial effects in various diseases, herbal medicines have long been accepted as adjunct therapeutics in cancer treatment. The growing body of evidence reveals that berberine-BBR (chemical formula: C₂₀H₁₈NO₄, slowly soluble in water), as an isoquinoline alkaloid found in numerous types of medicinal plants such as *Coptidis rhizoma*, *Phellodendron chinense schneid*, and *Phellodendron amurense*, is able to promote apoptosis in several types of tumors through the inhibition of IAPs family and the activation of caspase-9 and mitochondrial-mediated pathways (11, 12). Berberine may downregulate the expression of X-linked inhibitor of apoptotic proteins (XIAP), leading to the induction of apoptosis in leukocyte-depleted p53 genes. Moreover, berberine hydrochloride plays a focal role in reactive oxygen species generation, mitochondrial function, DNA topoisomerase inhibition, DNA or RNA binding, matrix metalloproteinase regulation, p53 activation, and NF-kappa B signal activation. Recently, the anti-metastatic and anti-cancer features of berberine against several cancers, including lung, prostate, breast, and gastric cancer, have been reported. In particular, the anti-proliferative and pro-apoptotic impacts of berberine have been demonstrated on leukemic cells derived from CLL patients (13, 14).

2. Objectives

Although studies have focused on the apoptotic role of berberine on CLL cells through mitochondrial pathway and the suppression of anti-apoptotic Bcl-2 gene expression, its impact on Apollon remains largely unknown. This study aimed to investigate the impact of berberine on the levels of Apollon gene expression in CLL patients and healthy donors.

3. Methods

3.1. Subjects

Totally, 12 patients newly-diagnosed with CLL and 6 healthy age-matched subjects who had no leukemia or other hematologic malignancies were enrolled in this study. The inclusion criteria for the diagnosis of CLL were

according to the International Workshop on Chronic Lymphocytic Leukemia guidelines (15). The main demographic features of the patients are summarized in Table 1. A total volume of 10 mL peripheral whole blood was obtained from the participants after written informed consent was obtained from them. The study was approved by the Ethics Committee of Semnan University of Medical Sciences (IR.SEMUMS.REC.1394.174).

Table 1. Demographic and Clinical Features of Chronic Lymphocytic Leukemia Patients

Patient No.	Gender	Age	WBC count/ μ L
1	M	65	59×10^3
2	M	71	84×10^3
3	M	74	65×10^3
4	F	52	62×10^3
5	F	68	48×10^3
6	M	42	53×10^3
7	M	50	43×10^3
8	F	64	84×10^3
9	F	69	85×10^3
10	M	64	62×10^3
11	F	66	73×10^3
12	M	72	71×10^3

Abbreviations: M, male; F, female.

3.2. Peripheral Blood Mononuclear Cell Isolation and Cell Culture

Peripheral blood mononuclear cells (PBMCs) were isolated from fresh heparinized samples by Ficoll-Hypaque density gradient (Amersham Pharmacia Biotech, Sweden). Briefly, peripheral blood samples were layered over the Ficoll-Hypaque gradient. The tubes were centrifuged for 20 min at $490 \times g$, and isolated PBMCs were distributed at approximately 2×10^6 cells/well in 6-well plate in RPMI-1640 medium with 5% fetal bovine serum (FBS) and 1% penicillin/streptomycin (Gibco, USA) at 37°C in a humidified incubator with 5% CO₂ for 48 h in the two groups of treated with berberine (CAS number: 633-65-8; Sigma-Aldrich, USA) at 25 μ M concentration (according to the previous studies (16, 17) and untreated (as control group).

3.3. RNA Extraction, cDNA Synthesis, and qRT-PCR

Treated and untreated PBMCs were lysed in TRIzol (Invitrogen, USA) for total RNA extraction according to the manufacturer's instruction. The quality and quantity of the extracted RNA were assessed by gel electrophoresis

and Nano-Drop instrument (Thermo Scientific, USA), respectively. Ten micrograms of total RNA were reverse transcribed to cDNA using the high capacity cDNA reverse kit (Thermo Scientific, USA). Real time q-PCR amplification was performed using the SYBR Green Master Mix (Amplicon, Denmark) using the real-time PCR detection system (Applied Biosystems, USA). Moreover, GAPDH was used as the housekeeping internal control gene. Primer sequences are listed in Table 2.

Table 2. The Primer Sequences of Apollon and GAPDH Genes

Gene	Sequences
Apollon	
Forward	5'-AGTGCAACGATGTGCCAT-3'
Reverse	5'-GCTAACCAACAGAGAGTA-3'
GAPDH	
Forward	5'-CCGTCTAGAAAACCTGCC-3'
Reverse	5'-AGCCAAATTCGTTGCATACC-3'

3.4. Statistical Analysis

All the statistical calculations were conducted with Prism version 7.03 (GraphPad Software). After assessment of normality, un-paired and paired t-test was performed to detect the significant differences between CLL patients and healthy donors and between berberine-treated and untreated groups, respectively.

4. Results

4.1. Classification of the Patients and Their Characteristics

A total of 12 patients and 6 age-matched healthy donors were enrolled in the study. The mean age of the patients was 63.8 years (range: 42 - 74 years). Moreover, all the patients are classified as stage 0 (only lymphocytosis) according to the Rai classification of CLL (18).

4.2. The Expression Levels of Apollon in CLL

The expression levels of Apollon gene were assessed in PBMCs of CLL patients and compared with that of adult healthy donors using qRT-PCR. No significant difference was identified in the Apollon expression level of CLL patients compared with the healthy donors ($P = 0.640$; Figure 1).

4.3. The Impact of Berberine on the Level of Apollon Expression in CLL

To assess the effects of berberine on the level of Apollon expression in PBMCs of CLL patients, the mean mRNA expression levels of Apollon was compared to that in the control groups by qRT-PCR. Comparative analysis revealed no significant differences between the berberine-treated group compared with the untreated group, both in CLL patients ($P = 0.545$; Figure 2A) and healthy donors ($P = 0.267$; Figure 2B), respectively.

5. Discussion

In this study, we addressed the in vitro effects of berberine on the levels of Apollon gene expression in PBMCs of CLL patients. Although our results revealed that the expression level of Apollon was not significantly different in CLL patients as compared to healthy donors, its tendency to decrease was clear in CLL patients. Indeed, Apollon is one of the eight members of IAP family that is heavily involved in the cell cycle regulation and DNA damage repair. The evasion of apoptosis through the dysregulation of IAPs has been reported to contribute to poor progression (10). The overexpression of Apollon has been observed in melanoma, prostate cancer, non-small cell lung cancer, and colorectal cancer (19).

Chomik et al. have shown that the up-regulation of Apollon gene expression is seen in advanced stages of CLL and is related to chemoresistance (20), whereas, in the current study, all the CLL patients were stratified in the stage 0 of Rai system. Therefore, it seems that the downregulated expression of Apollon in CLL patients is related to their disease stage. However, both upregulation and downregulation of Apollon gene expression were observed in different tumor types.

Despite the low expression of Apollon in childhood AML, which was confirmed by Schl  fli et al. (21), its upregulation is associated with unfavorable clinical outcomes and the high-proliferative feature of leukemic blasts (9). Berberine, as an isoquinoline quaternary alkaloid, exerts its anti-tumor activity in various human cancers mainly through interference with DNA replication, induction of the G1 phase cell cycle arrest, activation of autophagy, inhibition of tumor metastasis, and angiogenesis (22). This herbal derivative appears to be able to promote apoptosis through the inhibition of IAPs. In this context, Pandey et al. demonstrated that berberine can target survivin and STAT3 in gastric cancer cells (23). In this study, we assessed the effect of berberine on the level of Apollon expression in CLL patients and healthy donors. We found no significant differences between berberine-treated cells and untreated cells, both in CLL patients and healthy donors.

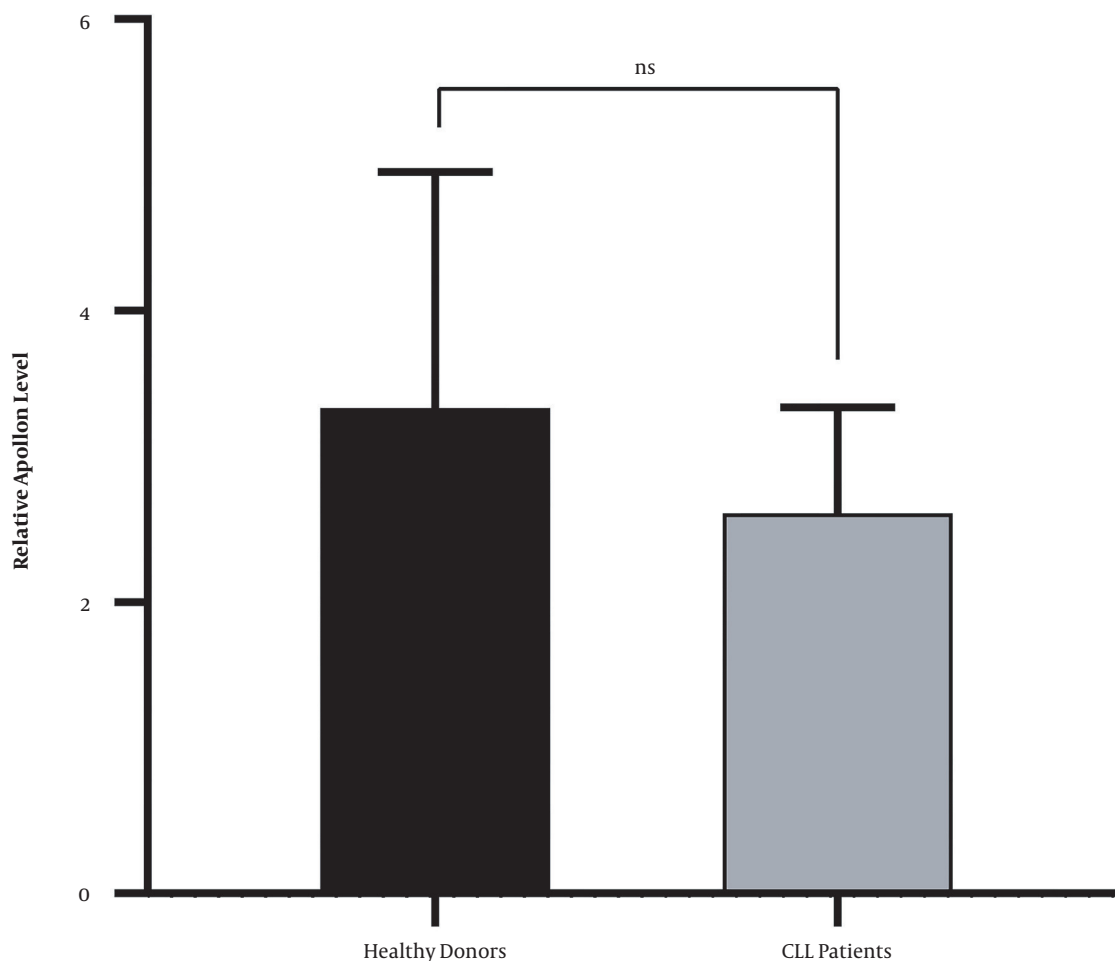


Figure 1. The expression level of Apollon in CLL patients and healthy donors. The Apollon levels of peripheral blood mononuclear cells of chronic lymphocytic leukemia (CLL) patients and healthy donors were measured using qRT-PCR. No significant difference was identified in the Apollon expression levels of CLL patients compared with the healthy donors ($P = 0.640$). Data are presented as mean \pm SEM (ns = no significant).

One of the hallmarks of CLL is the overexpression of anti-apoptotic Bcl-2 protein (24). However, berberine may trigger the intrinsic pathway of apoptosis through the downregulation of Bcl-2 by releasing cytochrome c in the mitochondrial membrane, and subsequently, the activation of caspase-9 besides activating caspases-8 and caspase-9 (25, 26). Moreover, based on the previous studies, berberine is able to significantly decrease Bcl-2/Bax ratio and up-regulate Fas/Fas ligand and p53 in tumor cells (27). This suggests that the ability of berberine to induce apoptosis in CLL is primarily dependent on the reduction of Bcl-2 expression, not Apollon.

5.1. Conclusion

In sum, our findings revealed that berberine has no significant impact on the expression of Apollon gene in CLL patients compared with healthy donors. However, further investigations are warranted to assess the exact effect of berberine on other IAPs.

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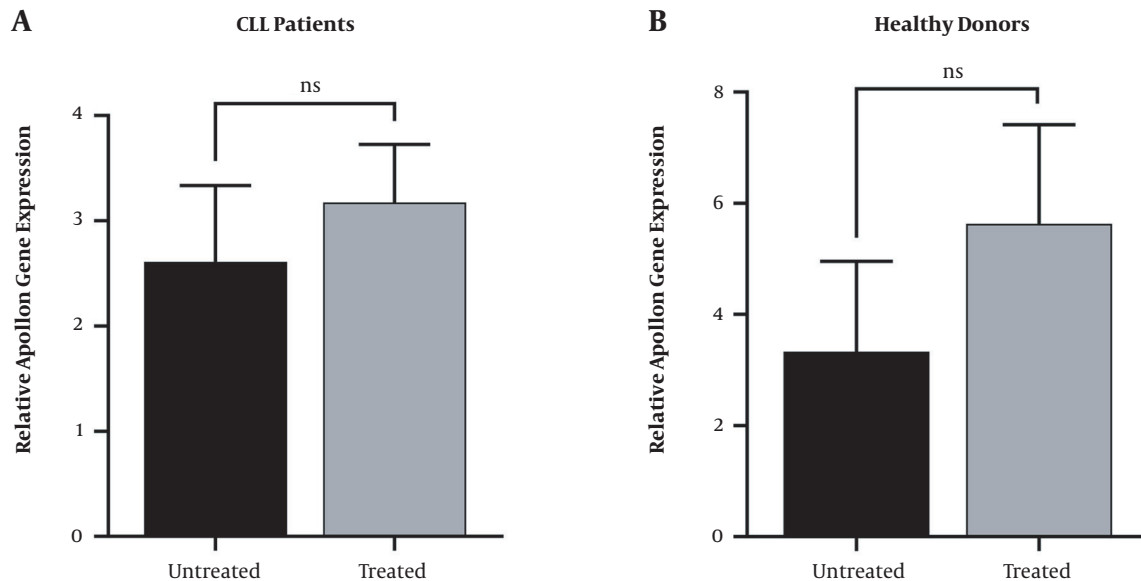


Figure 2. Apollon gene expression upon treatment with berberine in chronic lymphocytic leukemia patients and healthy donors peripheral blood mononuclear cells were cultured by 25 μ M berberine treatment for 48 hours compared with the untreated (control) groups. Subsequently, the Apollon levels were measured using qRT-PCR. Comparative analysis revealed no significant differences between berberine-treated groups and untreated groups, both in chronic lymphocytic leukemia patients ($P = 0.545$, Figure 2A) and healthy donors ($P = 0.267$; Figure 2B), respectively. Data are presented as mean \pm SEM (ns= no significant).

Footnotes

Authors' Contribution: Study concept and design, P.K; Acquisition of data, H. J and M.P; Analysis and interpretation of data, N.G and M. H; Drafting of the manuscript, N.G and M. H; Critical revision of the manuscript for important intellectual content, P.K; Statistical analysis, N.G and M. H; Administrative, technical, and material support, P.K; Study supervision, P.K.

Conflict of Interests: None to declare.

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Informed Consent: Written informed consent was obtained.

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